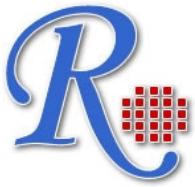
RayBio®
Human I-TAC/CXCL11
ELISA Kit

User Manual
(Revised Mar 1, 2012)

RayBio® Human I-TAC/CXCL11 ELISA Kit Protocol

(Cat#: ELH-ITAC-001)



RayBiotech, Inc.

We Provide You With Excellent
Protein Array System And Service

Tel:(Toll Free)1-888-494-8555 or 770-729-2992; Fax:770-206-2393;
Web: www.raybiotech.com Email: info@raybiotech.com

RayBiotech, Inc.

RayBio® Human I-TAC ELISA Kit Protocol

TABLE OF CONTENTS

I. INTRODUCTION

I-TAC (Interferon-inducible T-cell alpha chemoattractant) belongs to the group of CXC-Chemokines, was identified through sequence analysis of cDNAs derived from primary human astrocytes activated by cytokines. I-TAC expression is regulated by IFN. Human neutrophils produce I-TAC in response to IFN-gamma in combination with either TNF-alpha or bacterial lipopolysaccharides and this response is blocked by IL10 and IL4. IFN-gamma, alone or in association with agonists such as fMLP, IL8, G-CSF and GM-CSF have no effect. The factor is a potent chemoattractant for T-cells activated by IL2, but not for freshly isolated unstimulated T-cells, neutrophils, or monocytes. The receptor for I-TAC is CXCR3, which also functions as a receptor for IP-10 and human MIG.

The RayBio® Human I-TAC ELISA (Enzyme-Linked Immunosorbent Assay) kit is an in vitro enzyme-linked immunosorbent assay for the quantitative measurement of human I-TAC in serum, plasma (heparin or EDTA), cell culture supernatants and urine. This assay employs an antibody specific for human I-TAC coated on a 96-well plate. Standards and samples are pipetted into the wells and I-TAC present in a sample is bound to the wells by the immobilized antibody. The wells are washed and biotinylated anti-human I-TAC antibody is added. After washing away unbound biotinylated antibody, HRP-conjugated streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of I-TAC bound. The stop solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

II. REAGENTS

1. I-TAC Microplate (Item A): 96 wells (12 strips x 8 wells) coated with anti-human I-TAC.
2. Wash Buffer Concentrate (20x) (Item B): 25 ml of 20x concentrated solution.

3. Standards (Item C): 2 vials, recombinant human I-TAC.
4. Assay Diluent A (Item D): 30 ml of animal serum with 0.09% sodium azide as preservative. For Standard/Sample (serum/plasma) diluent.
5. Assay Diluent B (Item E): 15 ml of 5x concentrated buffer. For Standard/Sample (cell culture medium/urine) diluent.
6. Detection Antibody I-TAC (Item F): 2 vial of biotinylated anti-human I-TAC (each vial is enough to assay half microplate).
7. HRP-Streptavidin Concentrate (Item G): 200 µl of 200x concentrated HRP-conjugated streptavidin.
8. TMB One-Step Substrate Reagent (Item H): 12 ml of 3,3',5,5'-tetramethylbenzidine (TMB) in buffered solution.
9. Stop Solution (Item I): 8 ml of 0.2 M sulfuric acid.

III. STORAGE

May be stored for up to 6 months at 2° to 8°C from the date of shipment. Standard (recombinant protein) should be stored at -20 °C or -80 °C (recommended at –80 °C) after reconstitution. Opened Microplate Wells or reagents may be store for up to 1 month at 2° to 8°C. Return unused wells to the pouch containing desiccant pack, reseal along entire edge.
Note: the kit can be used within one year if the whole kit is stored at -20 °C. Avoid repeated freeze-thaw cycles.

IV. ADDITIONAL MATERIALS REQUIRED

1 Microplate reader capable of measuring absorbance at 450 nm.
2 Precision pipettes to deliver 2 µl to 1 ml volumes.
3 Adjustable 1-25 ml pipettes for reagent preparation.
4 100 ml and 1 liter graduated cylinders.
5 Absorbent paper.
6 Distilled or deionized water.
7 Log-log graph paper or computer and software for ELISA data analysis.

8 Tubes to prepare standard or sample dilutions.

V. REAGENT PREPARATION

1. Bring all reagents and samples to room temperature (18 - 25°C) before use.

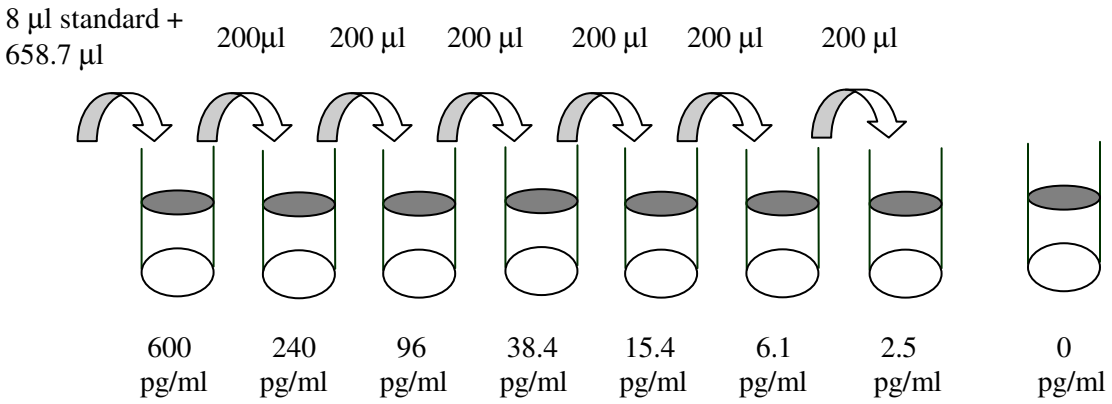
2. Sample dilution: If your samples need to be diluted, Assay Diluent A (Item D) should be used for dilution of serum/plasma samples. 1x Assay Diluent B (Item E) should be used for dilution of culture supernates and urine.

   Suggested dilution for normal serum/plasma: 2-5 fold*.

   *Please note that levels of the target protein may vary between different specimens. Optimal dilution factors for each sample must be determined by the investigator.

3. Assay Diluent B should be diluted 5-fold with deionized or distilled water.

4. Preparation of standard: **Briefly spin the vial of Item C** and then add 400 µl Assay Diluent A (for serum/plasma samples) or 1x Assay Diluent B (for cell culture medium and urine, Assay Diluent B should be diluted 5-fold with deionized or distilled water) into Item C vial to prepare a 50 ng/ml standard. **Dissolve the powder thoroughly by a gentle mix.** Add 8 µl I-TAC standard from the vial of Item C, into a tube with 658.7 µl Assay Diluent A or 1x Assay Diluent B to prepare a 600 pg/ml stock standard solution. Pipette 300 µl Assay Diluent A or 1x Assay Diluent B into each tube. Use the stock standard solution to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. Assay Diluent A or 1x Assay Diluent B serves as the zero standard (0 pg/ml).



5. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1x Wash Buffer.

6. Briefly spin the Detection Antibody vial (Item F) before use. Add 100 µl of 1x Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the concentrate can be stored at 4°C for 5 days). The detection antibody concentrate should be diluted 80-fold with 1x Assay Diluent B and used in step 4 of Part VI Assay Procedure.

7. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use. HRP-Streptavidin concentrate should be diluted 200-fold with 1x Assay Diluent B.

   For example: Briefly spin the vial (Item G) and pipette up and down to mix gently . A*dd 60 µl of HRP-Streptavidin concentrate into a tube with 12 ml 1x Assay Diluent B to prepare a final 200 fold diluted HRP- Streptavidin solution (don't store the diluted solution for next day use). Mix well.*

VI. ASSAY PROCEDURE:

1. Bring all reagents and samples to room temperature (18 - 25°C) before use. It is recommended that all standards and samples be run at least in duplicate.

2. Add 100 µl of each standard (see Reagent Preparation step 2) and sample into appropriate wells. Cover well and incubate for 2.5 hours at room temperature or over night at 4°C with gentle shaking.

3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel Pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

4. Add 100 µl of 1x prepared biotinylated antibody (Reagent Preparation step 6) to each well. Incubate for 1 hour at room temperature with gentle shaking.

5. Discard the solution. Repeat the wash as in step 3.

6. Add 100 µl of prepared Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking.

7. Discard the solution. Repeat the wash as in step 3.

8. Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking.

9. Add 50 µl of Stop Solution (Item I) to each well. Read at 450 nm immediately.

VII. ASSAY PROCEDURE SUMMARY

1. Prepare all reagents, samples and standards as instructed.

⇩

2. Add 100 µl standard or sample to each well.
   Incubate 2.5 hours at room temperature or over night at 4°C.

⇩

3. Add 100 µl prepared biotin antibody to each well.
   Incubate 1 hour at room temperature.

⇩

4. Add 100 µl prepared Streptavidin solution.
   Incubate 45 minutes at room temperature.

⇩

5. Add 100 µl TMB One-Step Substrate Reagent to each well.
   Incubate 30 minutes at room temperature.

⇩

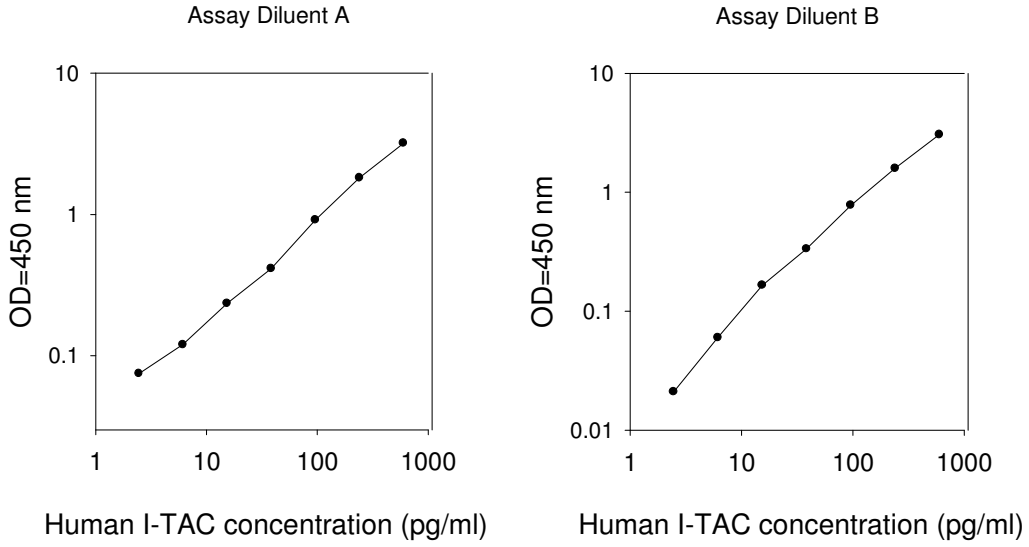
6. Add 50 µl Stop Solution to each well.
   Read at 450 nm immediately.

VIII. CALCULATION OF RESULTS

Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the average zero standard optical density. Plot the standard curve on log-log graph paper or using Sigma plot software, with standard concentration on the x-axis and absorbance on the y-axis. Draw the best-fit straight line through the standard points.

A. TYPICAL DATA

These standard curves are for demonstration only. A standard curve must be run with each assay.



B. SENSITIVITY

The minimum detectable dose of I-TAC is typically less than 2 pg/ml.

C. RECOVERY

Recovery was determined by spiking various levels of human I-TAC into human serum, plasma and cell culture media. Mean recoveries are as follows:

Sample Type	Average % Recovery	Range (%)
Serum	95.28	84-106
Plasma	93.47	83-104
Cell culture media	92.82	80-105

D. LINEARITY

Sample Type		Serum	Plasma	Cell Culture Media
1:2	Average % of Expected	94	93	90
	Range (%)	82-103	81-102	80-101
1:4	Average % of Expected	95	96	93
	Range (%)	83-106	82-105	82-104

E. REPRODUCIBILITY

Intra-Assay: CV<10%
Inter-Assay: CV<12%

IX. SPECIFICITY

Cross Reactivity: This ELISA kit shows no cross-reactivity with any of the cytokines tested *(e.g.*, human Angiogenin, BDNF, BLC, ENA-78, FGF-4, IL-1α, IL-1β, IL-2, IL-3, IL-4, IL-5, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12 p70, IL-12 p40, IL-13, IL-15, IL-309, IP-10, G-CSF, GM-CSF, IFN-γ, Leptin, MCP-1, MCP-2, MCP-3, MDC, MIP-1α, MIP-1 β, MIP-1δ, PARC, PDGF, RANTES, SCF, TARC, TGF-β, TIMP-1, TIMP-2, TNF-α, TNF-β, TPO, VEGF).

X. REFERENCES:

1. Gasperini S et al Gene expression and production of the monokine induced by IFN-gamma (mig), IFN-inducible T cell alpha chemoattractant (I-TAC), and IFN-gamma-inducible protein-10 (IP-10) chemokines by human neutrophils. Journal of Immunology 162(8): 4928-4937 (1999)

2. Cole KE et al Interferon-inducible T cell alpha chemoattractant (I-TAC): a novel non-ELR CXC chemokine with potent activity on activated T cells through selective high affinity binding to CXCR3. Journal of Experimental Medicine 187(12): 2009-2021 (1998)

3. Marx N et al Peroxisome proliferator-activated receptor-gamma Activators inhibit IFN-gamma-induced expression of the T cell-active CXC chemokines IP-10, mig, and I-TAC in human endothelial cells. Journal of Immunology 164(12): 6503-6508 (2000)

XI. TROUBLESHOOTING GUIDE

Problem	Cause	Solution
1. Poor standard curve	1. Inaccurate pipetting	1. Check pipettes
	2. Improper standard dilution	2. Ensure a brief spin of Item C and dissolve the powder thoroughly by a gentle mix.
2. Low signal	1.Too brief incubation times	1. Ensure sufficient incubation time; assay procedure step 2 may change to over night
	2. Inadequate reagent volumes or improper dilution	2. Check pipettes and ensure correct preparation
3. Large CV	1. Inaccurate pipetting	1. Check pipettes
4. High background	1. Plate is insufficiently washed	1. Review the manual for proper wash. If using a plate washer, check that all ports are unobstructed.
	2. Contaminated wash buffer	2. Make fresh wash buffer
5. Low sensitivity	1. Improper storage of the ELISA kit	1. Store your standard at<-20^{o}C after reconstitution, others at 4 ^{o}C. Keep substrate solution protected from light
	2. Stop solution	2. Stop solution should be added to each well before measure

RayBio® ELISA kits:

Over 200 ELISA kits, custom ELISA kit choose from over 500 list visit www.raybiotech.com for details.

This product is for research use only.